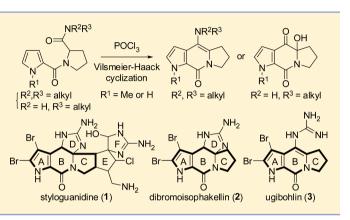
Synthesis and Unexpected Oxidization of the Tricyclic Core of Ugibohlin, Isophakellin, and Styloguanidine

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Supporting Information

ABSTRACT: A series of 4-substituted 5,6,7,9-tetrahydropyrrolo[2,3-*f*]indolizin-9-ones, representing the tricyclic core skeleton of ugibohlin, isophakellin, and styloguanidine, were synthesized via an intramolecular Vilsmeier—Haack reaction. This reaction allows the chemoselective C–C bond formation between the pyrrole C3 and proline C5 of N-[(pyrrol-2yl)carbonyl]prolinamides to construct the B-ring without the protection of the pyrrole nitrogen. Unexpected oxidizative property of the tricyclic core skeleton was observed, which could illuminate understanding of the biological formation of these marine secondary metabolites.



INTRODUCTION

A variety of structurally novel secondary metabolites from marine sponges have been identified to contain pyrrole and pyrrolidine ring units.¹ The most intriguing molecules in this family are the isomeric hexacyclic palau'amine^{2,3} and styloguanidine^{3,4} (1). Their A–B–C–D ring units are identical to dibromophakellin⁵ and dibromoisophakellin (cantharelline)⁶ (2), respectively. In addition, ugibohlin⁷ (3) is the D-ring seco-isomer of dibromoisophakellin (2) with an aromatized B-ring (Figure 1). These marine alkaloids exhibit interesting biological

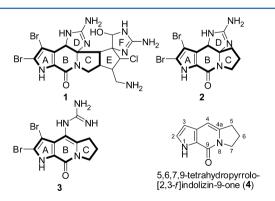


Figure 1. Marine natural products possessing tetrahydropyrrolo[2,3-f]indolizin-9-one core structure.

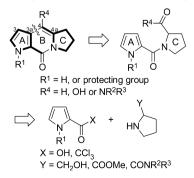
properties such as antitumor activities, substantial immunosuppressive activities, and low toxicities.⁸ The biological profiles combined with their characteristic structural motifs have made these alkaloids attractive targets for total synthesis.

To date, the total synthesis of phakellin has been accomplished by several research groups.^{9,10} Even the

challenging synthesis of palau'amine has recently been disclosed in the literature.¹¹ Nevertheless, the synthesis toward ugibohlin¹² (3) and dibromoisophakellin^{10,12} (2) was achieved only from the chemical conversion of the presynthesized dibromophakellin. Moreover, the total synthesis of styloguanidine (1) remains intangible. Since ugibohlin (3), dibromoisophakellin (2) and styloguanidine (1) all possess the same tricyclic core structure (Figure 1), the formation of the tricyclic A-B-C ring skeleton could be a common approach for the synthesis of these marine natural products.

The tricyclic core structure simply consists of pyrrole-2carboxylic acid and proline skeletons as ring A and C counterparts, respectively. The retrosynthetic analysis, delineated in Scheme 1, suggested that the construction of the tricyclic core could be attained by the *N*-acylation of the proline derivatives with pyrrole-2-carboxylic acid followed by the C–C

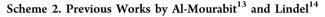
Scheme 1. Retrosynthetic Analysis of the Tricyclic Core

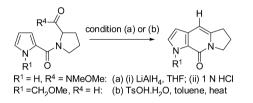


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bond formation between pyrrole ring-carbon (C3) and proline carbonyl carbon (C5). Although numerous methods have been developed, the electrophilic aromatic substitution is still one of the most promising reactions for the C–C bond formation at pyrrole ring carbons. Hence, the acid-promoted electrophilic aromatic substitution reaction, Friedel–Crafts acylation, and Vilsmeier–Haack reaction should be the most applicable methods for the preparation of various tricyclic A–B–C ring precursors.

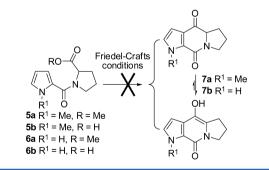
From a literature survey, Al-Mourabit¹³ and Lindel¹⁴ both demonstrated that the acid-promoted electrophilic aromatic substitution of N-[(pyrrol-2-yl)carbonyl]pyrrolidine-2-carbalde-hydes effectively afforded the tricyclic pyrrolo[2,3-*f*]indolizine skeleton (Scheme 2). But their further elaboration at the C4-and C4a-positions of the pyrroloindolizine achieved only limited success.





Our study focused on the construction of the tricyclic core and the introduction of functionality at the 4-position on the B ring. In our initial investigation, we recognized that the 4hydroxy-tricyclic A–B–C ring (7a,b) could be presented as its tautomeric keto form, and retrosynthetic disconnection between C-3a and C-4 would provide N-[(pyrrol-2-yl)carbonyl]proline derivatives (5a,b, 6a,b) as the ring A–C adducts (Scheme 3). The C–C bond formation between the

Scheme 3. Unsuccessful Attempts by Friedel–Crafts Cyclization

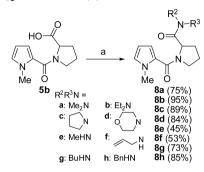


pyrrole C3 and the proline C5 was originally anticipated to be accomplished by Friedel–Crafts acylation reaction. However, attempts to promote the intramolecular Friedel–Crafts acylation of the ring A–C adducts (**5a,b, 6a,b**)^{13,15,16} by various Lewis acids or activating agents were all unsuccessful, which prompted us to investigate other strategies for the ring construction. We report herein the concise synthesis of the tricyclic core structure from pyrrole-2-carboxylic acid and proline via the Vilsmeier–Haack reaction as the key-step reaction and the unexpected oxidizative property of the pyrrolo[2,3-c]pyridin-7-one (the A–B ring).

RESULTS AND DISCUSSIONS

The N-[(pyrrol-2-yl)carbonyl]prolinamide substrates for the intramolecular Vilsmeier—Haack reaction were prepared by two different routes. N-[(1-Methylpyrrol-2-yl)carbonyl]proline¹⁶ (**5b**), readily prepared from commercially available 1-methylpyrrole and proline, as the acid component was reacted with a series of primary and secondary amines in the presence of EDC and HOBt to afford the corresponding N-[(1-methylpyrrol-2-yl)carbonyl]prolinamides (**8a**-**h**) (Scheme 4).

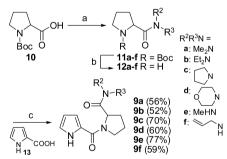
Scheme 4. Preparation of N-[(1-Methylpyrrol-2-yl)carbonyl]prolinamides (8)^{*a*}



^{*a*}Reagents and conditions: (a) R²R³NH, EDC, HOBt, Et₃N, CH₂Cl₂, rt.

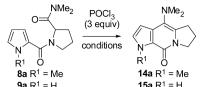
The same approach, however, failed to yield the pyrrole-*N*-unsubstituted *N*-[(pyrrol-2-yl)carbonyl]prolinamides (9a-f). Alternatively, a series of prolinamides (12a-f), prepared from *N*-Boc-proline (10), were coupled with pyrrole-2-carboxylic acid (13) to form the desired pyrrole-*N*-unsubstituted *N*-[(pyrrol-2-yl)carbonyl]prolinamides (9a-f) (Scheme 5).

Scheme 5. Preparation of N-[(Pyrrol-2-yl)carbonyl]prolinamides (9)^{*a*}



"Reagents and conditions: (a) R^2R^3NH , EDC, HOBt, Et₃N, CH₂Cl₂, 0 °C to rt; (b) aq. HCl/1,4-dioxane, rt; (c) EDC, HOBt, Et₃N, CH₂Cl₂, 0 °C to rt.

In the initial trials, N-[(1-methylpyrrol-2-yl)carbonyl]proline dimethylamide (8a) was chosen as a model compound to explore the reaction conditions for the Vilsmeier–Haack cyclization. Using a conventional approach, the dimethylamide 8a was reacted with phosphorus oxychloride in acetonitrile at 65 °C, and the reaction proceeded to give 31% of the product 14a, which was identified as the desired tricyclic adduct (entry 6 in Table 1). The preliminary success has confirmed that the Vilsmeier–Haack reaction is amenable for the construction of the tetrahydropyrrolo[2,3-f]indolizin-9-one ring with a dimethylamino substituent at the C4-position. Subsequently, various Table 1. Optimization for the Vilsmeier-Haack Cyclization



		9a R' = H	15a R' = H			
entry	reactant	base (2 equiv)	solvent	temperature (°C)	$_{(h)^a}^{time}$	yield (%) ^b
1	8a		CH_2Cl_2	reflux	24	39
2	8a	NMM	CH_2Cl_2	reflux	24	21
3	8a		THF	reflux	8	59
4	8a	NMM	THF	reflux	8	15
5	8a		CH ₃ CN	rt	36	4
6	8a		CH ₃ CN	65	2	31
7	8a	K ₂ CO ₃	CH ₃ CN	65	2	N.D. ^c
8	8a	DIPEA	CH ₃ CN	65	3	11
9	8a	DBU	CH ₃ CN	65	3	47
10	8a	Et_3N	CH ₃ CN	65	3	59
11	8a	NMM	CH ₃ CN	65	2	68
12	8a	NMM	CH ₃ CN	45	24	71
13	8a	NMM	CH ₃ CN	reflux	2	57
14	9a		CH_2Cl_2	reflux	3	39
15	9a		THF	rt	24	36
16	9a		THF	reflux	3	58
17	9a	NMM	THF	reflux	3	N.D. ^c
18	9a		DME	85	3	39
19	9a		CH ₃ CN	45	3	35
20	9a		CH ₃ CN	65	3	53
21	9a	NMM	CH ₃ CN	65	3	N.D. ^c

^{*a*}The reaction time was determined by TLC analysis based on the complete consumption of the reactant (8a, 9a). ^{*b*}Isolated yields. ^{*c*}N.D. = no desired product.

conditions including different base, solvent, temperature, time and the amount of the reagents were screened in order to optimize the cyclization reaction (Table 1). The optimum condition was to carry out the reaction with 3 equiv of POCl₃ and 2 equiv of *N*-methylmorpholine (NMM) in CH₃CN at 65 °C, and the reaction could be completed within 2 h (entry 11 in Table 1). However, when this condition was applied to the pyrrole-*N*-unsubstituted *N*-[(pyrrol-2-yl)carbonyl]proline dimethylamide (**9a**), the reaction did not result in any desired product. Thus, the reaction conditions were screened again, and the best cyclization condition for **9a** was to carry out the reaction with 3 equiv of POCl₃ in THF at reflux for 3 h (entry 16 in Table 1). It is noteworthy that the ring closure took place chemoselectively at the C3-position of pyrrole (C3–C5' bond formation) regardless the pyrrole nitrogen has a substituent.

With the optimized conditions in hand, a series of N-[(pyrrol-2-yl)carbonyl]proline dialkylamides (8a-d, 9a-d) were subjected to the Vilsmeier—Haack cyclization conditions, and the corresponding 4-dialkylamino-substituted 5,6,7,9-tetrahydropyrrolo[2,3-f]indolizin-9-ones (14a-d, 15a-d) were obtained in good yields (Table 2). When the Vilsmeier—Haack cyclization conditions were applied to N-[(pyrrol-2-yl)carbonyl]proline monoalkyl amides (8e-h, 9e,f), the initially formed products from individual reactions were observed on the TLC plates after workup. However, attempts to purify these products by flash column chromatography were unsuccessful, but all the initial products from individual reactions were converted to the same oxidized products, α -

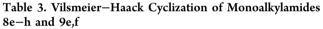
$\begin{array}{c} R^2 \\ N-R^3 \\ O \\ R^1 \\$										
entry	reactant	R¹	NR ² R ³	condition ^a	time (h)	product (yield, %) ^c				
1 ^{<i>d</i>}	8a	Me	NMe_2	А	2	14a (68)				
2	8b	Me	NEt_2	A	3	14b (48)				
3	8c	Me	N	А	10	14c (34)				
4	8d	Me	N_O	А	7	14d (62)				
5°	9a	Н	NMe_2	В	3	15a (58)				
6	9b	Н	NEt_2	В	3	15b (47)				
7	9c	Н	N	В	3	15c (44)				
8	9d	Н	NO	В	18	15d (13)				

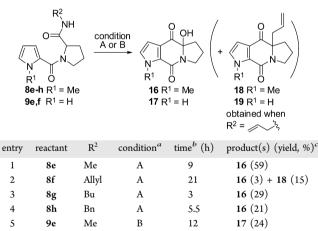
Table 2. Vilsmeier-Haack Cyclization of Dialkylamides 8a-

d and 9a-d

^{*a*}Condition A: POCl₃ (3 equiv), NMM (2 equiv), CH₃CN, 65 °C. Condition B: POCl₃ (3 equiv), THF, reflux. ^{*b*}The reaction time was determined by TLC analysis based on the complete consumption of the reactant. ^{*c*}Isolated yields. ^{*d*}Table 1, entry 11. ^{*e*}Table 1, entry 16.

hydroxy ketones (16, 17^{13} in Table 3), which were purified and characterized by spectroscopic analysis. The α -hydroxy ketones





"Condition A: POCl₃ (3 equiv), NMM (2 equiv), CH₃CN, 65 °C. Condition B: POCl₃ (3 equiv), THF, reflux. ^bThe reaction time was determined by TLC analysis based on the complete consumption of the reactant. 'Isolated yields.

14

17 (21) + 19 (7)

В

(16, 17) have a higher oxidation state than the natural products at C4-carbon. Attempts to reduce the α -hydroxy ketones (16, 17) to the dihydroxyl functionality were unsuccessful.

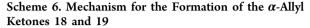
The conversion of the initial products, which might be the desired cyclized products, into the oxidized adducts likely involved an autoxidation after the Vilsmeier—Haack cyclization. The unexpected oxidation prompted us to examine the reaction more closely. When *N*-allylamides of *N*-[(pyrrol-2-yl)-carbonyl]prolines (**8f**, **9f**) were subjected to the Vilsmeier—Haack cyclization conditions, 4a-allyl-4,4a,5,6,7,9-hexahydropyrrolo[2,3-f]indolizine-4,9-diones (**18**, **19**) were obtained

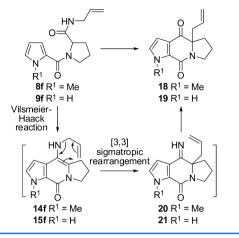
6

9f

Allyl

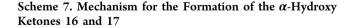
accompanied with the α -hydroxy ketones (16, 17). We rationalized that the α -allyl ketone byproducts (18, 19) were resulted from the aza-Claisen rearrangement of the allylamino adducts (14f, 15f) followed by hydrolysis of the imine intermediates (20, 21). This sequence of putative reactions provided evidence for the formation of the desired products from the Vilsmeier–Haack cyclization (Scheme 6).

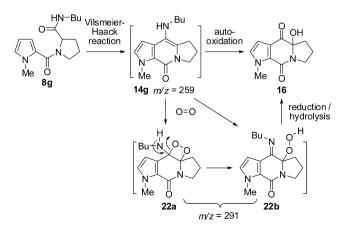




Meanwhile, the crude mixture obtained from the reaction of N-[(1-methylpyrrol-2-yl)carbonyl]proline butylamide (8g) after workup was immediately subjected to mass spectrometric analysis in order to gain insight into the reaction details. The molecular ion (M^+ , m/z = 259) for the expected Vilsmeier– Haack cyclization adduct was observed if the crude mixture was immediately preserved under argon. Upon exposure to air for an extended time, the appearance of the predominant M+32 ion (m/z = 291), corresponding to a hydroperoxide (22a) or a dioxetane (22b) intermediate, reflected the immediate autoxidation of the Vilsmeier–Haack cyclization product. The hydroperoxide 22b was degraded in the slightly acidic silica gel column to form the oxidized product 16 (Scheme 7).

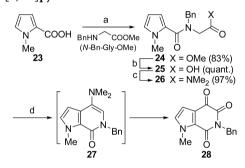
We postulated that the unexpected oxidization was attributed to the oxidizable tendency of the A–B ring unit. Thus, we embarked on the synthesis of the A–B ring unit in order to test whether the amino-substituted pyrrolo[2,3-c]pyridine-7-one would be aerobically oxidizable. Analogous to the tricyclic





system, *N*-methylpyrrole-2-carboxylic acid (23) was coupled with *N*-benzylglycine methyl ester, and the resulting ester 24 was saponified followed by coupling with dimethylamine hydrochloride to afford *N*-benzyl-*N*-[(pyrrol-2-yl)carbonyl]glycine dimethylamide (26). The dimethylamide substrate 26 was subjected to the Vilsmeier–Haack cyclization condition, and the ¹H NMR analysis suggested that the initial product obtained from the reaction immediately after column chromatography was 1-methyl-4-dimethylamino-6-benzylpyrrolo[2,3-c]pyridin-7-one (27). However, upon exposure to the air for a short period of time, the cyclized adduct 27 was oxidized into the corresponding pyrrolo[2,3-c]pyridine-4,5,7trione 28,¹⁷ the structure of which was confirmed by X-ray crystallographic analysis (Scheme 8). It is notable that even

Scheme 8. Synthesis and Oxidation of 4-Amino-Substituted Pyrrolo[2,3-c]pyridin-7-one^a



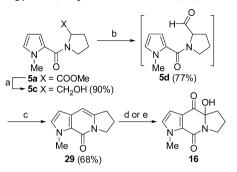
^aReagents and conditions: (a) EDC, HOBt, Et_3N , CH_2Cl_2 , rt, 18 h; (b) (i) aq. NaOH/MeOH, reflux, 3 h; (ii) HCl; (c) Me_2NH.HCl, EDC, HOBt, Et_3N , CH_2Cl_2 , rt, 18 h; (d) POCl_3, NMM, CH_3CN , 65 °C, 20 h, 37% for **28** (from **26**).

though the tricyclic 4-diakylamino-substituted tetrahydropyrrolo[2,3-*f*]indolizin-9-ones (14a-d, 15a-d) were stable enough to be purified and characterized, a gradual oxidative degradation of 14a-d and 15a-d into 16 and 17, respectively, and other unidentified decomposition products was observed in a prolonged time period.

To further clarify the role of the 4-amino-substituents in the autoxidation, the 4-unsubstituted tetrahydropyrrolo [2,3-f]indolizin-9-one 29 was prepared by adopting Lindel's approach.¹⁴ N-[(1-Methylpyrrol-2-yl)carbonyl]proline methyl ester 5a was first reduced by lithium borohydride and then oxidized by IBX¹⁸ to form the corresponding aldehyde 5d. Subsequently, the acid-promoted electrophilic ring-closure afforded the 4-unsubstituted tricyclic core skeleton 29. Unlike the 4-amino-substituted congeners, the 4-unsubstituted tricyclic core 29 shows substantial stability under aerobic conditions. Therefore, the electron-donating character of the 4-aminosubstituents is responsible for the unexpected aerobic oxidation of the pyrrolo[2,3-c]pyridin-7-one heterocycle. Furthermore, attempts to carry out the dihydroxylation and epoxidation¹³ at the C4-C4a double bond of the tricyclic core 29 were unsuccessful, but interestingly, both resulted in the formation of the α -hydroxyketone adduct 16 (Scheme 9).

CONCLUSION

In summary, we have developed a facile and practical synthesis for the tricyclic core skeleton, tetrahydropyrrolo[2,3-f]indolizin-9-one, of ugibohlin, isophakellin, and styloguanidine via an intramolecular Vilsmeier—Haack reaction. Our investigation also revealed that the nitrogen-substituent at the 4Scheme 9. Synthesis and Oxidation of the 4-Unsubstituted Tetrahydropyrrolo [2,3-f] indolizin-9-one $(29)^a$



^aReagents and conditions: (a) NaBH₄/LiCl, THF/EtOH, 0 °C to rt, 8 h; (b) IBX, EtOAc, reflux, 3 h; (c) MsOH, CH_2Cl_2 , rt, 1.5 h; (d) cat. OsO₄, NMO, acetone, rt, 3 h, 58%; (e) *m*CPBA, CH_2Cl_2 , 0 °C to rt, 2 h, 63%.

position of the tricyclic core provoked the aerobic oxidation of the heterocycle. The highly oxidizable tendency of the pyrrolo[2,3-c]pyridine-7-one ring unit (A–B ring) could illuminate understanding of the biological formation of these marine secondary metabolites and also present a challenge for the total synthesis of these marine natural products.

EXPERIMENTAL SECTION

General Chemical Procedures. The chemical shift values are reported in δ values (parts per million, ppm) relative to the standard chemical shift for the hydrogen residue peak and carbon-13 peak in the deuterated solvent, $CDCl_3$, or DMSO- d_6 .¹⁹ The coupling constant (J) values are expressed in hertz (Hz). The numbers of protons directly attached to the individual carbons were determined by ¹³C NMR DEPT experiments. High-resolution mass spectrometry (HRMS) was obtained by the following ionization methods/mass analyzers: EI/ magnetic sector, ESI/TOF or FAB/magnetic sector. Thin-layer chromatography (TLC) was performed on silica gel plates. Compounds on TLC were visualized by illumination under UV light (254 nm), or using the following reagents: phosphomolybdic acid reagent, anisaldehyde reagent, ninhydrin reagent or 10% ethanolic sulfuric acid followed by charring on a hot plate. Silica gel (230-400 mesh) was used for flash column chromatography, and this technique has been described by Still et al.²⁰ Evaporations were carried out under reduced pressure (water aspirator or vacuum pump) with the bath temperature below 50 °C unless specified otherwise. Materials obtained from commercial suppliers were used without further purification.

N-[(1-Methylpyrrol-2-yl)carbonyl]proline dimethylamide (8a). A mixture of N-[(1-methylpyrrol-2-yl)carbonyl]proline¹⁰ (5b)(4.0869 g, 18.39 mmol), EDC (5.1816 g, 27.03 mmol, 1.5 equiv) and HOBt (3.6397 g, 26.94 mmol, 1.5 equiv) in CH₂Cl₂ (100 mL) was stirred for 30 min at room temperature under argon. Dimethylammonium chloride (2.9351 g, 36.0 mmol, 2 equiv) and triethylamine (12.6 mL, 89.65 mmol, 5 equiv) were added to the solution. The reaction mixture was stirred for an additional 6 h at ambient temperature under argon and then concentrated under reduced pressure. The residue was dissolved in CHCl₃ (50 mL), washed with 1 N aqueous HCl solution, saturated aqueous Na2CO3 solution, and saturated aqueous NaCl solution. The organic layer was dried over anhydrous MgSO4 and concentrated under reduced pressure. The residue was purified by flash column chromatography (Hex/EtOAc = 1:9, $R_f = 0.1$) to give 8a (amorphous solid, 3.37 g, 13.5 mmol, 75%): mp 125-128 °C; ¹H NMR (CDCl₃, 400 MHz) δ 1.84–1.89 (m, 2 H), 1.92–2.24 (m, 2 H), 2.95 (bs, 3 H), 3.15 (bs, 3 H), 3.81 (bs, 3 H), 3.85 (bs, 2 H), 4.99 (bs, 1 H), 6.05 (bs, 1 H), 6.59 (bs, 1 H), 6.64 (bs, 1 H); ¹³C NMR (CDCl₃, 100 MHz) & 25.4 (CH₂), 28.5 (CH₂), 35.9 (CH₃), 36.5 (CH₃), 37.0 (CH₃), 50.0 (CH₂), 56.8 (CH), 106.7 (CH), 114.1 (CH),

125.7, 126.7 (CH), 161.7, 172.0; MS (EI, 20 eV) m/z 108 (52), 177 (100), 249 (20) (M⁺); HRMS (EI, magnetic sector) calcd for C₁₃H₁₉N₃O₂ 249.1477, found 249.1485.

N-[(1-Methylpyrrol-2-yl)carbonyl]proline diethylamide (8b). Compound **8b** was prepared from **5b** by the method described for **8a**. The reaction was purified by flash column chromatography (Hex/ EtOAc = 4:6, $R_f = 0.1$) to give **8b** (oil, 1.0546 g, 95%): ¹H NMR (CDCl₃, 400 MHz) δ 0.83–0.95 (m, 3 H), 1.07–1.25 (m, 3 H), 1.84– 1.90 (m, 2 H), 2.02–2.08 (m, 1 H), 2.10–2.18(m, 1 H), 3.12–3.23 (m, 1 H), 3.23–3.26 (m, 1 H), 3.31–3.33 (m, 1 H), 3.43–3.56 (m, 1 H), 3.76–3.81 (m, 5 H), 4.91 (bs, 1 H), 6.01 (bs, 1 H), 6.53 (bs, 1 H), 6.60 (bs, 1 H); ¹³C NMR (CDCl₃, 100 MHz) δ 12.8 (CH₃), 14.5 (CH₃), 25.3 (CH₂), 29.2 (CH₂), 36.1 (CH₃), 40.6 (CH₂), 41.8 (CH₂), 50.0 (CH₂), 56.5 (CH), 106.6 (CH), 113.7 (CH), 125.8, 126.3 (CH), 161.7, 171.5; MS (EI, 20 eV) *m*/*z* 108 (50), 177 (100), 277 (17) (M⁺); HRMS (EI, magnetic sector) calcd for C₁₅H₂₃N₃O₂ 277.1790, found 277.1797.

N-[(1-Methylpyrrol-2-yl)carbonyl]proline (pyrrolidin-1-yl)amide (8c). Compound 8c was prepared from 5b by the method described for 8a. The reaction was purified by flash column chromatography (EtOAc, $R_f = 0.1$) to give 8c (colorless oil, 0.5623 g, 89%): ¹H NMR (CDCl₃, 400 MHz) δ 1.76–1.89 (m, 6 H), 2.07– 2.14 (m, 2 H), 3.35–3.37 (m, 2 H), 3.47–3.49 (m, 1 H), 3.75–3.78 (m, 9 H), 4.73 (bs, 1 H), 5.99 (bs, 1 H), 6.52 (bs, 1 H), 6.58 (bs, 1 H); ¹³C NMR (CDCl₃, 100 MHz) δ 23.8 (CH₂), 25.3 (CH₂), 25.9 (CH₂), 28.2 (CH₂), 36.2 (CH₃), 45.7 (CH₂), 45.9 (CH₂), 49.9 (CH), 58.1 (CH₂), 106.4 (CH), 113.8 (CH), 125.5, 126.4 (CH), 161.5, 170.3; MS (EI, 20 eV) *m*/*z* 108 (100), 177 (70), 275 (11) (M⁺); HRMS (EI, magnetic sector) calcd for C₁₅H₂₁N₃O₂ 275.1634, found 275.1642.

N-[(1-Methylpyrrol-2-yl)carbonyl]proline (morpholin-1-yl)amide (8d). Compound **8d** was prepared from **5b** by the method described for **8a**. The reaction was purified by flash column chromatography (EtOAc, $R_f = 0.2$) to give **8d** (oil, 0.3902 g, 84%): ¹H NMR (CDCl₃, 400 MHz) δ 1.84–1.91 (m, 2 H), 2.09–2.21 (m, 2 H), 3.60–3.83 (m, 13 H), 4.95–4.97 (m, 1 H), 6.04 (bs, 1 H), 6.58 (bs, 1 H), 6.63 (bs, 1 H); ¹³C NMR (CDCl₃, 100 MHz) δ 25.4 (CH₂), 28.8 (CH₂), 36.3 (CH₃), 42.3 (CH₂), 46.0 (CH₂), 49.9 (CH₂), 56.4 (CH), 66.5 (CH₂), 66.7 (CH₂), 106.7 (CH), 114.1 (CH), 125.5, 126.8 (CH), 161.7, 170.7; MS (EI, 20 eV) *m/z* 108 (32), 177 (100), 291 (14) (M⁺); MS (EI, 70 eV) *m/z* 108 (33), 177 (100), 291 (13) (M⁺); HRMS (EI, magnetic sector) calcd for C₁₅H₂₁N₃O₃ 291.1583, found 291.1589.

N-[(1-Methylpyrrol-2-yl)carbonyl]proline methylamide (8e). Compound 8e was prepared from 5b by the method described for 8a. The reaction was purified by flash column chromatography (EtOAc, R_f = 0.1) to give 8e (amorphous solid, 1.4702 g, 45%): mp 114–117 °C; ¹H NMR (CDCl₃, 400 MHz) δ 1.84–1.92 (m, 1 H), 1.98–2.07 (m, 2 H), 2.30–2.36 (m, 1 H), 2.78 (s, 3 H), 3.66–3.74 (m, 1 H), 3.80–3.87 (m, 4 H), 4.71 (dd, 1 H, *J* = 4.9 and 7.3 Hz), 6.08 (dd, 1 H, *J* = 2.6 and 3.8 Hz), 6.54 (bs, 1 H), 6.71 (t, 1 H, *J* = 1.9 Hz); ¹³C NMR (CDCl₃, 100 MHz) δ 25.2, 26.2, 27.5, 36.7, 50.1, 60.3, 107.1, 114.6, 125.0, 127.6, 163.3, 172.2; MS (EI, 20 eV) *m*/*z* 108 (100), 177 (95), 235 (43) (M⁺); HRMS (EI, magnetic sector) calcd for C₁₂H₁₇N₃O₂ 235.1321, found 235.1321.

N-[(1-Methylpyrrol-2-yl)carbonyl]proline allylamide (8f). Compound 8f was prepared from **5b** by the method described for **8a**. The reaction was purified by flash column chromatography (Hex/ EtOAc = 2:8, R_f = 0.25) to give 8f (colorless oil, 0.69 g, 53%): ¹H NMR (CDCl₃, 400 MHz) δ 1.84–1.87 (m, 1 H), 1.97–2.04 (m, 2 H), 2.27–2.28 (m, 1 H), 3.66–3.74 (m, 1 H), 3.77–3.86 (m, 6 H), 4.71 (bs, 1 H), 5.02 (dd, 1 H, *J* = 1.2 and 10.3 Hz), 5.11 (d, 1 H, *J* = 17.1 Hz), 5.71–5.78 (m, 1 H), 6.04–6.05 (m, 1 H), 6.51 (bs, 1 H), 6.67 (bs, 1 H), 7.04 (bs, 1 H); ¹³C NMR (CDCl₃, 100 MHz) δ 25.3, 27.8, 36.7, 41.8, 50.0, 60.5, 107.1, 114.4, 115.9, 125.2, 127.5, 134.1, 163.2, 171.6; MS (EI, 20 eV) *m*/*z* 108 (42), 177 (100), 261 (33) (M⁺); HRMS (EI, magnetic sector) calcd for C₁₄H₁₉N₃O₂ 261.1477, found 261.1480. *N*-[(1-Methylpyrrol-2-yl)carbonyl]proline *n*-butylamide (8g). Compound 8g was prepared from 5b by the method described for 8a. The reaction was purified by flash column chromatography (Hex/ EtOAc = 7:3, $R_f = 0.1$) to give 8g (pale-yellow oil, 0.7280 g, 73%): ¹H NMR (CDCl₃, 400 MHz) δ 0.81 (t, 3 H, J = 7.4 Hz), 1.22–1.28 (m, 2 H), 1.38–1.42 (m, 2 H), 1.83–1.89 (m, 1 H), 1.97–2.02 (m, 2 H), 2.25–2.26 (m, 1 H), 3.15–3.19 (m, 2 H), 3.67–3.69 (m, 1 H), 3.77– 3.78 (m, 4 H), 4.64–4.66 (m, 1 H), 6.03–6.04 (m, 1 H), 6.49 (bs, 1 H), 6.66 (m, 1 H), 6.86; ¹³C NMR (CDCl₃, 100 MHz) δ 13.7, 20.0, 25.2 27.8, 31.5, 36.6, 39.2, 49.9, 60.7, 107.1, 114.4, 125.2, 127.4, 163.2, 171.7; MS (EI, 20 eV) m/z 108 (47), 177 (100), 277 (27) (M⁺); HRMS (EI, magnetic sector) calcd for C₁₅H₂₃N₃O₂ 277.1790, found 277.1801.

N-[(1-Methylpyrrol-2-yl)carbonyl]proline benzylamide (8h). Compound 8h was prepared from 5b by the method described for 8a. The reaction was purified by flash column chromatography (Hex/ EtOAc = 4:6, R_f = 0.2) to give 8h (brown amorphous solid, 1.4233 g, 85%): mp 108−111 °C; ¹H NMR (CDCl₃, 400 MHz) δ 1.87−1.93 (m, 1 H), 1.98−2.08 (m, 2 H), 2.33−2.36 (m, 1 H), 3.70−3.86 (m, 5 H), 4.44 (dd, 1 H, *J* = 5.9 and 15.0 Hz), 4.46 (dd, 1 H, *J* = 5.7 and 14.8 Hz), 4.76−4.78 (m, 1 H), 6.09 (bs, 1 H), 6.55 (bs, 1 H), 6.70 (bs, 1 H), 7.22−7.32 (m, 5 H); ¹³C NMR (CDCl₃, 100 MHz) δ 25.1 (CH₂), 27.9 (CH₂), 36.5 (CH₃), 43.2 (CH₂), 49.8 (CH₂), 60.5 (CH), 106.9 (CH), 114.3 (CH), 125.1, 127.1 (CH), 127.3 (CH), 128.4 (CH), 128.5 (CH), 138.3, 163.1, 171.6; MS (EI, 70 eV) *m/z* 108 (100), 109 (26), 177 (100), 178 (23), 311 (11) (M⁺); HRMS (EI, magnetic sector) calcd for C₁₈H₂₁N₃O₂ 311.1634, found 311.1643.

N-[(Pyrrol-2-yl)carbonyl]proline dimethylamide (9a). To a mixture of pyrrole-2-carboxylic acid (13, 1.67 g, 15.0 mmol), EDC (3.45 g, 18.0 mmol, 1.2 equiv) and HOBt (2.43 g, 18.0 mmol, 1.2 equiv) in CH_2Cl_2 (75 mL) was added proline dimethylamide hydrochloride salt²¹ (**12a**, obtained quantitatively from *N*-Boc-proline dimethylamide²² (11a) by acidic hydrolysis without further purification, 2.68 g, 15.0 mmol, 1.0 equiv) and triethylamine (10.5 mL, 75.0 mmol, 5.0 equiv) at 0 °C. The mixture was stirred at room temperature for 12 h. The solvent was removed under reduced pressure. The residue was dissolved in CH₂Cl₂ (100 mL), washed with H₂O and saturated aqueous NaCl solution. The organic layer was dried over anhydrous MgSO4 and concentrated under reduced pressure. The residue was purified by flash column chromatography (Hex/EtOAc = 7:3 to EtOAc) to give 9a (amorphous solid, 1.99 g, 8.46 mmol, 56%, $R_f = 0.10$ (EtOAc)): mp 182–183 °C; ¹H NMR (DMSO-d₆, 400 MHz) δ 1.72–1.76 (m, 1 H), 1.91–1.97 (m, 1 H), 2.00-2.05 (m, 1 H), 2.11-2.16 (m, 1 H), 2.81 (s, 3 H), 3.10 (s, 3 H), 3.76-3.81 (m, 2 H), 4.92-4.95 (m, 1 H), 6.15 (s, 1 H), 6.63 (s, 1 H), 6.90 (s, 1 H), 11.35 (bs, 1 H, NH); ¹³C NMR (DMSO-d₆, 100 MHz) δ 25.3 (CH₂), 28.3 (CH₂), 35.7 (CH₃), 37.1 (CH₃), 48.6 (CH₂), 57.6 (CH), 109.3 (CH), 112.5 (CH), 121.9 (CH), 126.0, 159.8, 172.0; MS (EI, 70 eV) m/z 66 (16), 70 (100), 94 (86), 163 (96), 235 (8) (M⁺); HRMS (EI, magnetic sector) calcd for C₁₂H₁₇N₃O₂ 235.1321, found 235.1325.

N-[(Pyrrol-2-yl)carbonyl]proline diethylamide (9b). Compound 9b was prepared from 12b^{21,23} and 13 by the method described for 9a. The reaction was purified by flash column chromatography (Hex/EtOAc = 4:6 to EtOAc) to give 9b (amorphous solid, 2.44 g, 52%, R_f = 0.30 (EtOAc)): mp 118–119 °C; ¹H NMR (CDCl₃, 400 MHz) δ 1.08 (t, 3 H, *J* = 7.2 Hz), 1.25 (t, 3 H, *J* = 7.2 Hz), 1.87–1.99 (m, 2 H), 3.23–3.38 (m, 2 H), 3.48–3.53 (m, 2 H), 3.85–3.97 (m, 2 H), 4.95–4.98 (m, 1 H), 6.17 (s, 1 H), 6.62 (s, 1 H), 6.84 (s, 1 H), 10.51 (bs, 1 H, NH); ¹³C NMR (CDCl₃, 100 MHz) δ 12.9 (CH₃), 14.4 (CH₃), 25.2 (CH₂), 28.8 (CH₂), 40.6 (CH₂), 41.7 (CH₂), 48.6 (CH₂), 57.6 (CH), 109.4 (CH), 112.8 (CH), 121.5 (CH), 125.3, 160.4, 171.4; MS (EI, 70 eV) *m*/*z* 66 (11), 70 (100), 94 (82), 163 (97), 263 (9) (M⁺); HRMS (EI, magnetic sector) calcd for C₁₄H₂₁N₃O₂ 263.1634, found 263.1638.

N-[(Pyrrol-2-yl)carbonyl]proline (pyrrolidin-1-yl)amide (9c). Compound 9c was prepared from $12c^{21,24}$ and 13 by the method described for 9a. The reaction was purified by flash column chromatography (Hex/EtOAc = 5:5 to EtOAc) to give 9c (amorphous solid, 1.76 g, 70%, $R_f = 0.10$ (EtOAc)): mp 194–195 °C; ¹H NMR (500 MHz, CDCl₃) δ 1.85–1.88 (m, 2 H), 1.96–2.06 (m, 4 H), 2.14–2.16 (m, 1 H), 2.32–2.34 (m, 1 H), 3.39–3.49 (m, 2 H), 3.58–3.63 (m, 1 H), 3.86–3.91 (m, 2 H), 4.02 (bs, 1 H), 4.82–4.85 (m, 1 H), 6.25 (d, 1 H, 2.2 Hz), 6.66 (s, 1 H), 6.91 (s, 1 H), 9.59 (bs, 1 H, NH); ¹³C NMR (125 MHz, CDCl₃) δ 24.1 (CH₂), 25.4 (CH₂), 26.3 (CH₂), 28.3 (CH₂), 46.0 (CH₂), 46.3 (CH₂), 48.5 (CH₂), 59.2 (CH), 109.9 (CH), 112.6 (CH), 121.1 (CH), 125.6, 160.3, 170.5; MS (EI, 70 eV) m/z 70 (100), 94 (69), 163 (87), 261 (21) (M⁺); HRMS (EI, magnetic sector) calcd for C₁₄H₁₉N₃O₂ 261.1477, found 261.1482.

N-[(Pyrrol-2-yl)carbonyl]proline (morpholin-1-yl)amide (9d). Compound 9d was prepared from $12d^{25,26}$ and 13 by the method described for 9a. The reaction was purified by flash column chromatography (Hex/EtOAc = 4:6 to EtOAc) to give 9d (amorphous solid, 1.95 g, 60%, R_f = 0.10 (EtOAc)): mp 184–185 °C; ¹H NMR (400 MHz, CDCl₃) δ 1.91–1.95 (m, 1 H), 1.99–2.07 (m, 1 H), 2.11–2.19 (m, 1 H), 2.26 (m, 1 H), 3.57–3.72 (m, 7 H), 3.79–3.81 (m, 1 H), 3.89–3.91 (m, 1 H), 3.99–4.00 (m, 1 H), 5.03–5.06 (m, 1 H), 6.24 (d, 1 H, *J* = 3.0 Hz), 6.66 (s, 1 H), 6.90 (s, 1 H), 9.91 (bs, 1 H, NH); ¹³C NMR (100 MHz, CDCl₃) δ 25.3 (CH₂), 28.5 (CH₂), 42.4 (CH₂), 46.1 (CH₂), 48.4 (CH₂), 57.2 (CH), 66.6 (CH₂), 66.9 (CH₂), 109.9 (CH), 112.7 (CH), 121.4 (CH), 125.4, 160.3, 170.6; MS (EI, 70 eV) *m/z* 66 (16), 70 (100), 94 (94), 163 (99), 164 (13), 277 (5) (M⁺); HRMS (EI, magnetic sector) calcd for C₁₄H₁₉N₃O₃ 277.1426, found 277.1433.

N-[(**Pyrrol-2-yl**)**carbonyl**]**proline methylamide (9e).** Compound 9e was prepared from $12e^{22,27}$ and 13 by the method described for 9a. The reaction was purified by flash column chromatography (Hex/EtOAc = 6:4 to EtOAc) to give 9e (amorphous solid, 1.93 g, 77%, $R_f = 0.13$ (EtOAc)): mp 112–113 °C; ¹H NMR (400 MHz, CDCl₃) δ 1.91–2.38 (m, 4 H), 2.74 (d, 3 H, J = 3.6 Hz), 3.75–3.85 (m, 2 H), 4.78 (d, 1 H, J = 4.0 Hz), 6.25 (s, 1 H), 6.66 (s, 1 H), 6.95 (s, 1 H), 7.05 (s, 1 H, NH), 10.29 (s, 1 H, NH); ¹³C NMR (100 MHz, CDCl₃) δ 25.5 (CH₂), 26.2 (CH₃), 27.2 (CH₂), 48.7 (CH₂), 61.1 (CH), 110.3 (CH), 113.3 (CH), 122.1 (CH), 125.0, 161.7, 172.0; MS (EI, 70 eV) m/z 66 (6), 70 (100), 94 (58), 163 (76), 221 (6) (M⁺); HRMS (EI, magnetic sector) calcd for C₁₁H₁₅N₃O₂ 221.1164, found 221.1164.

N-[(Pyrrol-2-yl)carbonyl]proline allylamide (9f). Compound 9f was prepared from 12f²⁸ and 13 by the method described for 9a. The reaction was purified by flash column chromatography (Hex/EtOAc = 7:3 to EtOAc) to give 9f (amorphous solid, 2.78 g, 59%, R_f = 0.33 (EtOAc)): mp 115–116 °C; ¹H NMR (400 MHz, CDCl₃) δ 1.94–2.44 (m, 4 H), 3.77–3.91 (m, 4 H), 4.83 (d, 1 H, *J* = 6.6 Hz), 5.06 (d, 1 H, *J* = 10.2 Hz), 5.15 (d, 1 H, *J* = 17.2 Hz), 5.79–5.80 (m, 1 H), 6.28 (s, 1 H), 6.68 (s, 1 H), 6.97 (s, 1 H), 7.20 (bs, 1 H, NH), 9.98 (bs, 1 H, NH); ¹³C NMR (100 MHz, CDCl₃) δ 25.5 (CH₂), 27.0 (CH₂), 41.7 (CH₂), 48.6 (CH₂), 61.2 (CH), 110.4 (CH), 113.3 (CH), 115.8 (CH₂), 121.9 (CH), 125.0, 134.1 (CH), 161.7, 171.2; MS (EI, 70 eV) *m*/*z* 66 (19), 70 (100), 94 (90), 163 (99), 164 (22), 247 (8) (M⁺); HRMS (EI, magnetic sector) calcd for C₁₃H₁₇N₃O₂ 247.1321, found 247.1320.

4-Dimethylamino-1-methyl-5,6,7,9-tetrahydropyrrolo[2,3*f*]indolizin-9-one (14a). To a solution of 8a (0.2501 g, 1.0 mmol) in acetonitrile (20 mL) at 0 $^\circ C$ was added phosphorus oxychloride (0.28 mL, 3.05 mmol, 3 equiv) and N-methylmorpholine (0.22 mL, 2.0 mmol, 2 equiv). After the addition was completed, the reaction mixture was heated at 65 $^{\circ}\mathrm{C}$ for 2 h. The solution was cooled to room temperature and then concentrated under reduced pressure. The residue was dissolved in chloroform (25 mL), washed with saturated aqueous Na2CO3 solution, saturated NaCl solution, dried over anhydrous MgSO4 and concentrated under reduced pressure. The resulting residue was purified by flash column chromatography (EtOAc, $R_f = 0.5$) to give 14a (pale-yellow amorphous solid, 0.1573) g, 0.68 mmol, 68%): mp 94–97 °C; ¹H NMR (CDCl₃, 500 MHz) δ 2.08–2.14 (m, 2 H), 2.76 (s, 6 H), 3.05 (t, 2 H, J = 7.2 Hz), 4.08 (t, 2 H, J = 7.3 Hz), 4.11 (s, 3 H), 6.31 (s, 1 H), 6.89 (s, 1 H); ¹³C NMR $(CDCl_3, 125 \text{ MHz}) \delta 22.4 (CH_2), 29.1 (CH_2), 35.5 (CH_3), 44.0$ (CH₃), 47.4 (CH₂), 100.3 (CH), 123.0, 123.3, 130.5 (CH), 130.9, 135.4, 154.0; MS (EI, 20 eV) m/z 108 (18), 177 (16), 216 (36), 231

(100) (M⁺); HRMS (EI, magnetic sector) calcd for $C_{13}H_{17}N_3O$ 231.1372, found 231.1372.

4-Diethylamino-1-methyl-5,6,7,9-tetrahydropyrrolo[**2**,**3**-*f*]**indolizin-9-one (14b).** Compound 14b was prepared from 8b by the method described for 14a. The reaction was purified by flash column chromatography (Hex/EtOAc = 1:9 then EtOAc) to give 14b (brown oil, 86.9 mg, 48%, R_f = 0.25 (EtOAc)): ¹H NMR (CDCl₃, 400 MHz) δ 0.89 (t, 6 H, *J* = 7.1 Hz), 2.04–2.11 (m, 2 H), 3.00–3.06 (m, 6 H), 4.07–4.10 (m, 5 H), 6.23 (d, 1 H, *J* = 2.4 Hz), 6.85 (d, 1 H, *J* = 2.2 Hz); ¹³C NMR (CDCl₃, 100 MHz) δ 14.0 (CH₃), 22.4 (CH₂), 29.7 (CH₂), 35.5 (CH₃), 47.8 (CH₂), 48.7 (CH₂), 100.5 (CH), 118.6, 123.0, 130.4 (CH), 131.5, 139.1, 154.1; MS (EI, 70 eV) *m/z* 187 (20), 188 (29), 229 (18), 230 (77), 244 (73), 259 (100) (M⁺); HRMS (EI, magnetic sector) calcd for C₁₅H₂₁N₃O 259.1685, found 259.1690.

1-Methyl-4-(pyrrolidin-1-yl)-5,6,7,9-tetrahydropyrrolo[2,3*f*]**indolizin-9-one (14c).** Compound 14c was prepared from 8c by the method described for 14a. The reaction was purified by flash column chromatography (EtOAc, $R_f = 0.2$) to give 14c (white amorphous solid, 81.4 mg, 34%): mp 101–104 °C; ¹H NMR (CDCl₃, 400 MHz) δ 1.92–1.95 (m, 4 H), 2.08–2.15 (m, 2 H), 3.06 (t, 2 H, *J* = 7.6 Hz), 3.14–3.17 (m, 4 H), 4.09 (t, 2 H, *J* = 7.3 Hz), 4.12 (s, 3 H), 6.21 (d, 1 H, *J* = 2.6 Hz), 6.89 (d, 1 H, *J* = 2.6 Hz); ¹³C NMR (CDCl₃, 100 MHz) δ 22.4 (CH₂), 25.6 (CH₂), 29.2 (CH₂), 29.6 (CH₂), 35.6 (CH₃), 47.6 (CH₂), 51.4 (CH₂), 100.0 (CH), 120.0, 123.1, 130.5 (CH), 130.8, 136.3, 153.9; MS (EI, 70 eV) *m*/*z* 256 (23), 257 (100) (M⁺); HRMS (EI, magnetic sector) calcd for C₁₅H₁₉N₃O 257.1528, found 257.1536.

1-Methyl-4-(morpholin-1-yl)-5,6,7,9-tetrahydropyrrolo[2,3*f*]**indolizin-9-one (14d).** Compound 14d was prepared from 8d by the method described for 14a. The reaction was purified by flash column chromatography (EtOAc, $R_f = 0.25$) to give 14d (brown amorphous solid, 0.1466 g, 62%): mp 148–151 °C; ¹H NMR (CDCl₃, 400 MHz) δ 2.10–2.17 (m, 2 H), 3.07–3.12 (m, 6 H), 3.78 (t, 4 H, *J* = 4.4 Hz), 4.01 (t, 2 H, *J* = 7.0 Hz), 4.13 (s, 3 H), 6.35 (d, 1 H, *J* = 2.8 Hz), 6.91 (d, 1 H, *J* = 2.8 Hz); ¹³C NMR (CDCl₃, 100 MHz) δ 22.4 (CH₂), 29.2 (CH₂), 35.5 (CH₃), 47.6 (CH₂), 51.5 (CH₂), 68.0 (CH₂), 100.3 (CH), 122.0, 122.9, 130.7 (CH), 131.2, 136.2, 154.0; MS (EI, 70 eV) *m*/*z* 108 (13), 214 (15), 215 (36), 273 (100) (M⁺); HRMS (EI, magnetic sector) calcd for C₁₅H₁₉N₃O₂ 273.1477, found 273.1480.

4-Dimethylamino-5,6,7,9-tetrahydropyrrolo[2,3-f]indolizin-**9-one (15a).** To a solution of **9a** (0.47 g, 2.0 mmol) in THF (20 mL) was added POCl₃ (0.55 mL, 6.0 mmol, 3.0 equiv) at 0 °C. After the addition was completed, the mixture was stirred at reflux temperature for 3 h. The solution was cooled to room temperature, and the solvent was removed under reduced pressure. The residue was partitioned between CHCl₃ and H₂O. The aqueous layer was separated and then neutralized with saturated aqueous Na2CO3 solution. The aqueous solution was extracted with CHCl₃, and the organic layer was dried over anhydrous MgSO₄ and concentrated under reduced pressure. The residue was purified by flash column chromatography (Hex/EtOAc = 2:8 to EtOAc) to give 15a (yellow amorphous solid, 0.1521 g, 0.7 mmol, 35%, $R_f = 0.25$ (EtOAc)): mp 219–220 °C; ¹H NMR (DMSO d_{6} , 500 MHz) δ 2.05–2.11 (m, 2 H), 2.72 (s, 6 H), 3.04 (t, 2 H, J = 6.0 Hz), 3.97 (t, 2 H, J = 5.7 Hz), 6.39 (t, 1 H, J = 1.6 Hz), 7.21 (t, 1 H, J = 2.0 Hz), 11.85 (bs, 1 H, NH); ¹³C NMR (DMSO- d_{61} 125 MHz) δ 22.1 (CH₂), 28.7 (CH₂), 43.9 (CH₃), 47.3 (CH₂), 101.5 (CH), 122.5, 123.5, 126.0 (CH), 129.1, 135.0, 152.4; MS (EI, 70 eV) m/z 202 (100), 217 (78) (M⁺); HRMS (EI, magnetic sector) calcd for C12H15N3O 217.1215, found 217.1219.

4-Diethylamino-5,6,7,9-tetrahydropyrrolo[**2,3-***f*]**indolizin-9-one** (15b). Compound 15b was prepared from 9b by the method described for 15a. The reaction was purified by flash column chromatography (Hex/EtOAc = 2:8 then EtOAc) to give 15b (yellow amorphous solid, 0.12 g, 21%, R_f = 0.18 (EtOAc)): mp 158–159 °C; ¹H NMR (CDCl₃, 400 MHz) δ 0.77–0.86 (m, 6 H), 2.05–2.11 (m, 2 H), 2.97–3.06 (m, 6 H), 3.23–3.38 (m, 2 H), 3.48–3.53 (m, 2 H), 3.85–3.97 (m, 6 H), 4.16 (t, 2 H, *J* = 7.2 Hz), 6.29 (s, 1 H), 7.17 (s, 1 H), 11.90 (bs, 1 H, NH); ¹³C NMR (CDCl₃, 100 MHz) δ 14.0 (CH₃), 22.6 (CH₂), 29.6 (CH₂), 29.7 (CH₂), 48.2 (CH₂), 48.9 (CH₂), 101.9 (CH), 119.8, 124.1, 126.6 (CH), 130.8, 138.7, 153.5; MS (EI, 70 eV)

m/z173 (34), 174 (24), 216 (58), 230 (90), 245 (100) (M⁺); HRMS (EI, magnetic sector) calcd for $\rm C_{14}H_{19}N_3O$ 245.1528, found 245.1530.

4-(Pyrrolidin-1-yl)-5,6,7,9-tetrahydropyrrolo[**2,3-f**]indolizin-**9-one (15c).** Compound **15c** was prepared from **9c** by the method described for **15a**. The reaction was purified by flash column chromatography (CH₂Cl₂/MeOH = 97:3) to give **15c** (amorphous solid, 0.21 g, 44%, R_f = 0.25 (EtOAc)): mp 225–226 °C; ¹H NMR (DMSO- d_{60} 500 MHz) δ 1.90 (bs, 4 H), 2.04–2.10 (m, 2 H), 3.02 (t, 2 H, *J* = 6.0 Hz), 3.34 (s, 4 H), 3.98 (t, 2 H, *J* = 5.6 Hz), 6.29 (s, 1 H), 7.21 (s, 1 H), 11.88 (bs, 1 H, NH); ¹³C NMR (DMSO- d_{60} 125 MHz) δ 22.6 (CH₂), 25.6 (CH₂), 29.3 (CH₂), 47.9 (CH₂), 51.7 (CH₂), 101.7 (CH), 119.5, 124.1, 126.5 (CH), 129.3, 136.2, 152.9; MS (EI, 70 eV) *m*/*z* 70 (93), 94 (100), 120 (34), 205 (35), 243 (44) (M⁺); HRMS (EI, magnetic sector) calcd for C₁₄H₁₇N₃O 243.1372, found 243.1371.

4-(Morpholin-1-yl)-5,6,7,9-tetrahydropyrrolo[2,3-*f*]indolizin-9-one (15d). Compound 15d was prepared from 9d by the method described for 15a. The reaction was purified by flash column chromatography (CH₂Cl₂/MeOH = 99:1 to 98:2) to give 15d (amorphous solid, 0.10 g, 13%, R_f = 0.1 (CH₂Cl₂/MeOH = 98/2)): mp 198–199 °C; ¹H NMR (DMSO- d_{64} 400 MHz) δ 2.05–2.12 (m, 2 H), 3.00 (s, 4 H), 3.08 (t, 2 H, *J* = 7.6 Hz), 3.69 (s, 4 H), 3.98 (t, 2 H, *J* = 7.0 Hz), 6.40 (s, 1 H), 7.22 (s, 1 H), 11.89 (bs, 1 H, NH); ¹³C NMR (DMSO- d_{64} 100 MHz) δ 22.6 (CH₂), 29.2 (CH₂), 47.9 (CH₂), 51.8 (CH₂), 67.7 (CH₂), 102.0, 121.7, 123.9, 126.7, 129.8, 136.0, 152.9; MS (EI, 70 eV) *m*/*z* 173 (14), 177 (10), 200 (15), 201 (47), 259 (100) (M⁺); HRMS (EI, magnetic sector) calcd for C₁₄H₁₇N₃O₂ 259.1321, found 259.1322.

1-Methyl-4a-hydroxy-4,4a,5,6,7,9-hexahydropyrolo[**2,3-f**]**indolizine-4,9-dione (16)**. *Method A*. To a solution of 8e (0.2340 g, 1.0 mmol) in acetonitrile (20 mL) at 0 °C was added phosphorus oxychloride (0.28 mL, 3.05 mmol, 3 equiv) and *N*-methylmorpholine (0.22 mL, 1.92 mmol, 2 equiv). After the addition was completed, the reaction mixture was heated at 65 °C for 16 h under argon. The reaction mixture was cooled to room temperature and then concentrated under reduced pressure. The residue was dissolved in chloroform (10 mL), washed with saturated aqueous Na₂CO₃ solution, saturated aqueous NaCl solution, dried over anhydrous MgSO₄ and concentrated under reduced pressure. The resulting residue was purified by flash column chromatography (Hex/EtOAc = 5:5, $R_f = 0.1$) to give **16** (amorphous solid, 0.1299 g, 0.59 mmol, 59%). The product was recrystallized from toluene to give an analytical sample of **16**: mp 136–137 °C (white solid, toluene).

Method B. To a solution of 29 (188 mg, 1.0 mmol) in acetone (1 mL) was added N-methylmorpholine N-oxide (NMO, 234.3 mg, 2.0 mmol, 2 equiv) and OsO_4 (25.4 mg, 0.10 mmol, 0.1 equiv). The reaction mixture was stirred at room temperature for 3 h. The reaction mixture was quenched with saturated aqueous Na₂SO₃ solution at 0 °C for 10 min. The solvent was removed under reduced pressure. The residue was dissolved in EtOAc (50 mL), washed with H₂O, saturated aqueous Na₂CO₃ solution, saturated aqueous NaCl solution, and then dried over anhydrous Na₂SO₄. The solvent was evaporated under reduced pressure. The residue was purified by flash column chromatography to give **16** (amorphous solid, 127.7 mg, 0.58 mmol, 58%).

Method C.¹³ To a solution of **29** (150 mg, 0.80 mmol) in CH₂Cl₂ (1 mL) was added *m*-chloroperbenzoic acid (*m*CPBA, 300 mg, 1.72 mmol, 2.2 equiv) at 0 °C. After the addition was completed, the reaction temperature was raised to room temperature, and the reaction mixture was stirred for 2 h. The mixiture was quenched with saturated aqueous Na₂SO₃ solution at 0 °C for 10 min. The residue was extracted with EtOAc, and the organic layer was washed with H₂O, saturated aqueous NaCl solution, dried over anhydrous Na₂SO₄ and then concentrated under reduced pressure. The residue was purified by flash column chromatography to give **16** (amorphous solid, 109.8 mg, 0.50 mmol, 63%): ¹H NMR (400 MHz, DMSO-*d*₆) δ 1.88–1.98 (m, 2 H), 2.02–2.12 (m, 2 H), 3.57–3.58 (m, 2 H), 3.94 (s, 3 H), 6.46 (d, 1 H, *J* = 2.6 Hz), 6.61 (s, 1 H), 7.15 (d, 1 H, *J* = 2.6 Hz); ¹³C NMR (100 MHz, DMSO-*d*₆) δ 20.1 (CH₂), 34.9 (CH₂), 36.0 (CH₃), 44.7 (CH₂), 91.4, 105.8 (CH), 122.7, 130.2, 130.9 (CH), 156.5, 188.7; MS

(EI, 20 eV) m/z 108 (100), 202 (27), 220 (50) (M⁺); HRMS (EI, magnetic sector) calcd for $C_{11}H_{12}N_2O_3$ 220.0848, found 220.0849. Anal. Calcd for $C_{11}H_{12}N_2O_3$: C, 59.99; H, 5.49; N, 12.72. Found: C, 59.96; H, 5.50; N, 12.51.

1-Methyl-4a-allyl-4,4a,5,6,7,9-hexahydropyrolo[2,3-f]indolizine-4,9-dione (18). To a solution of 8f (0.7727 g, 2.95 mmol) in acetonitrile (60 mL) at 0 °C was added phosphorus oxychloride (0.81 mL, 8.85 mmol, 3 equiv) and N-methylmorpholine (0.67 mL, 5.86 mmol, 2 equiv). After the addition was completed, the reaction mixture was heated at 65 °C for 21 h under argon. The reaction mixture was cooled to room temperature and then concentrated under reduced pressure. The residue was dissolved in chloroform (50 mL), washed with saturated aqueous Na2CO3 solution, saturated aqueous NaCl solution, dried over anhydrous MgSO4 and concentrated under reduced pressure. The resulting residue was purified by flash column chromatography (Hex/EtOAc = 5:5 to EtOAc) to give products 18 (amorphous solid, 0.1096 g, 0.45 mmol, 15%, $R_f = 0.5$ (EtOAc)) and 16 (amorphous solid, 0.0213 g, 0.10 mmol, 3%, $R_f = 0.3$ (EtOAc). 18: ¹H NMR (CDCl₃, 500 MHz) δ 1.95–2.03 (m, 3 H), 2.14–2.17 (m, 1 H), 2.45 (ddd, 1 H, J = 5.8, 10.9, and 16.7 Hz), 3.49-3.55 (m, 1 H), 3.85-3.90 (m, 1 H), 3.97 (s, 3 H), 4.92 (t, 2 H, J = 9.3 Hz), 5.41-5.50 (m, 1 H), 6.44 (d, 1 H, J = 2.9 Hz), 6.69 (d, 1 H, J = 2.9 Hz); ¹³C NMR (CDCl₃, 125 MHz) δ 20.5 (CH₂), 32.8 (CH₂), 35.9 (CH₃), 43.5 (CH₂), 44.1 (CH₂), 75.2, 105.1 (CH), 119.2, 124.1, 129.6 (CH), 131.2, 131.4 (CH), 156.8, 192.8; MS (EI, 20 eV) m/z 108 (15), 136 (92), 202 (77), 204 (100), 244 (35) (M⁺); HRMS (EI, magnetic sector) calcd for C14H16N2O2 244.1212, found 244.1212.

4a-Hydroxy-4,4a,5,6,7,9-hexahydropyrrolo[2,3-f]indolizine-**4,9-dione**¹³ (17). To a solution of 9e (0.2210 g, 1.00 mmol) in THF (10 mL) was added POCl₃ (0.28 mL, 3.0 mmol, 3.0 equiv) at 0 °C. After the addition was completed, the reaction mixture was heated at reflux temperature for 16 h. The reaction mixture was cooled to room temperature and then concentrated under reduced pressure. The residue was dissolved in CHCl₃/EtOH (15 mL/15 mL), and then Na₂CO₃ (3.18 g, 30 mmol, 30 equiv) was added. The mixture was stirred for 2.5 h. After filtration, the solvent was evaporated and the residue was purified by flash column chromatography (CH2Cl2/ MeOH = 99:1 to 98:2) to give the 17 (amorphous solid, 49.4 mg, 24%, $R_f = 0.28$ (EtOAc)): mp 188–189 °C;¹³ ¹H NMR (DMSO- d_{60} 400 MHz) δ 1.88–1.96 (m, 2 H), 2.06–2.12 (m, 2 H), 3.57–3.61 (m, 2 H), 6.49 (t, 1 H, J = 2.4 Hz), 6.62 (s, 1 H, OH), 7.10 (t, 1 H, J = 2.6 Hz), 12.60 (bs, 1 H, NH); $^{13}\mathrm{C}$ NMR (DMSO- $d_{6\prime}$ 100 MHz) δ 20.2 (CH₂), 34.8 (CH₂), 44.5 (CH₂), 91.8, 106.7 (CH), 121.7, 125.3 (CH), 132.7, 156.0, 188.9; MS (EI, 70 eV) m/z 93 (74), 94 (100), 206 (29) (M⁺); HRMS (EI, magnetic sector) calcd for $C_{10}H_{10}N_2O_3$ 206.0691, found 206.0694.

4a-Allyl-4,4a,5,6,7,9-hexahydropyrolo[2,3-f]indolizine-4,9dione (19). To a solution of 9f (5.40 g, 21.84 mmol) in THF (218 mL) was added POCl₃ (6.01 mL, 65.5 mmol, 3.0 equiv) at 0 °C. After the addition was completed, the reaction mixture was heated at reflux temperature for 14 h. The reaction mixture was cooled to room temperature and then concentrated under reduced pressure. The residue was dissolved in CH2Cl2. The solution was washed with saturated aqueous Na2CO3 solution, dried over anhydrous MgSO4 and concentrated under reduced pressure. The residue was purified by flash column chromatography ($Hex/CHCl_3 = 1:9$ then $CHCl_3/MeOH$ = 99:1) to give the products 19 (amorphous solid, 0.34 g, 1.48 mmol, 7%, $R_f = 0.45$ (EtOAc)) and 17 (amorphous solid, 0.95 g, 4.61 mmol, 21%, $R_f = 0.28$ (EtOAc)). 19: mp 138–139 °C; ¹H NMR (400 MHz, CDCl₃) δ 2.08–2.15 (m, 3 H), 2.22–2.28 (m, 1 H), 2.48–2.53 (m, 1 H), 2.59-2.64 (m, 1 H), 3.61-3.66 (m, 1 H), 3.99-4.06 (m, 1 H), 4.96-5.03 (m, 2 H), 5.47-5.57 (m, 1 H), 6.59 (s, 1 H), 7.02 (s, 1 H), 11.92 (bs, 1 H, NH); ¹³C NMR (100 MHz, CDCl₃) δ 20.5 (CH₂), 32.8 (CH₂), 43.5 (CH₂), 44.1 (CH₂), 75.7, 106.2 (CH), 119.4 (CH₂), 123.4, 125.1 (CH), 131.2 (CH), 133.0, 156.9, 192.8; MS (ESI) m/z 188 (34), 229 (100) (M - 1); HRMS (ESI, TOF) calcd for $C_{13}H_{13}N_2O_2$ 229.0977 (M - 1), found 229.0978.

N-[(1-Methylpyrrol-2-yl)carbonyl]-*N*-benzylglycine methyl ester (24). A mixture of *N*-methylpyrrole-2-carboxylic acid (23) (1.0572 g, 8.45 mmol, 1.2 equiv), EDC (1.6291 g, 8.50 mmol, 1.2 equiv) and HOBt (1.1482 g, 8.50 mmol, 1.2 equiv) in dichloromethane (50 mL) was stirred for 10 min at room temperature under argon. N-Benzylglycine methyl ester²⁹ (N-Bn-Gly-OMe, 1.2852 g, 7.17 mmol) and triethylamine (3.0 mL, 21.34 mmol, 3 equiv) were added to the solution, and the reaction mixture was stirred at room temperature for 18 h. The solution was concentrated under reduced pressure. The residue was dissolved in chloroform (30 mL), washed with H₂O and saturated aqueous NaCl solution, dried over anhydrous MgSO₄ and then concentrated under reduced pressure. The residue was purified by flash column chromatography (Hex/EtOAc = 9:1 then EtOAc) to give 24 (brown amorphous solid, 1.7011 g, 5.94 mmol, 83%, $R_f = 0.1$ (Hex/EtOAc = 9:1)): mp 91–94 °C; ¹H NMR (CDCl₃, 400 MHz) δ 3.72 (s, 3 H), 3.82 (s, 3 H), 4.12 (bs, 2 H), 4.89 (bs, 1 H), 6.00 (t, 2 H, J = 2.8 Hz), 6.36–6.37 (m, 1 H), 6.71 (s, 1 H), 7.25– 7.31 (m, 3 H), 7.35–7.39 (m, 2 H); ¹³C NMR (CDCl₃, 100 MHz) δ 36.0, 47.2, 52.1, 54.4, 107.0, 112.6, 124.3, 126.7, 127.6, 128.9, 136.7, 165.1, 169.9; MS (EI, 70 eV) m/z 91 (22), 108 (100), 178 (48), 286 (10) (M⁺); HRMS (EI, magnetic sector) calcd for C₁₆H₁₈N₂O₃ 286.1317, found 286.1322.

3-[[(1-Methylpyrrol-2-yl)carbonyl]-N-benzyl-amino]ethanic acid (25). To a solution of N-[(1-methylpyrrol-2-yl)carbonyl]-Nbenzylglycine methyl ester (24) (1.4493 g, 5.06 mmol) in methanol (30 mL) was added 15% aqueous NaOH solution (30 mL). The reaction mixture was heated at reflux temperature for 3 h. The mixture was concentrated under reduced pressure. The residue was partitioned between water and chloroform. The pH of the aqueous layer was adjusted to 2 with 4 N aqueous HCl solution. The aqueous solution was then extracted with chloroform (30 mL \times 3). The combined organic layer was dried over anhydrous ${\rm MgSO}_4$ and concentrated under reduced pressure to give 25, which was used for the subsequent reaction without further purification (brown oil, 1.3753 g, 5.05 mmol, quantitatively, $R_f = 0.5$ (Hex/EtOAc = 5:5)): ¹H NMR (CDCl₃, 400 MHz) δ 3.81 (s, 3 H), 4.14 (s, 2 H), 4.90 (bs, 2 H), 6.01 (t, 2 H, J = 2.8 Hz), 6.40 (t, 2 H, J = 1.0 Hz), 6.72 (s, 1 H), 7.26–7.40 (m, 5 H); $^{13}\mathrm{C}$ NMR (CDCl₃, 100 MHz) δ 36.1, 47.9, 54.2, 107.2, 113.2, 123.9, 127.0, 127.5, 127.7, 129.0, 136.3, 165.6, 173.4; MS (EI, 70 eV) m/z 81 (54), 91 (31), 108 (100), 109 (29), 272 (57) (M⁺); HRMS (EI, magnetic sector) calcd for C₁₅H₁₆N₂O₃ 272.1161, found 272.1163.

N-[(1-Methylpyrrol-2-yl)carbonyl]-N-benzylglycine dimethylamide (26). A mixture of N-[(1-methylpyrrol-2-yl)carbonyl]-Nbenzylglycine (25) (1.0106 g, 3.71 mmol), EDC (1.4180 g, 7.40 mmol, 2 equiv) and HOBt (1.0000 g, 7.40 mmol, 2 equiv) in CH₂Cl₂ (30 mL) was stirred for 10 min at room temperature under argon. Dimethylammonium chloride (0.4532 g, 5.55 mmol, 1.5 equiv) and triethylamine (2.1 mL, 14.80 mmol, 4 equiv) were added to the solution. The reaction mixture was stirred for an additional 19 h at ambient temperature under argon and then concentrated under reduced pressure. The residue was dissolved in chloroform (30 mL), washed with H2O and saturated aqueous NaCl solution, dried over anhydrous MgSO4 and then concentrated under reduced pressure. The residue was purified by flash column chromatography (Hex/ EtOAc = 3:7, $R_f = 0.1$) to give 26 (brown amorphous solid, 1.0744 g, 3.59 mmol, 97%): mp 92–95 °C; ¹H NMR (CDCl₃, 400 MHz) δ 2.90 (bs, 3 H), 2.95 (s, 3 H), 3.80 (s, 3 H), 4.16 (s, 2 H), 4.91 (bs, 2 H), 5.96 (bs, 1 H), 6.32 (bs, 1 H), 6.67 (bs, 1 H), 7.27-7.29 (m, 3 H), 7.34–7.37 (m, 2 H); ¹³C NMR (CDCl₃, 100 MHz) δ 35.6 (CH₃), 35.8 (CH₃), 36.1 (CH₃), 46.1 (CH₂), 54.0 (CH₂), 106.7 (CH), 111.8 (CH), 124.7, 126.4 (CH), 126.7 (CH), 127.3 (CH), 127.5 (CH), 128.7 (CH), 137.1, 165.2, 167.5; MS (EI, 70 eV) m/z 91 (34), 108 (100), 191 (95), 213 (71), 299 (4) (M⁺); HRMS (EI, magnetic sector) calcd for C₁₇H₂₁N₃O₂ 299.1634, found 299.1626.

6-Benzyl-1-methyl-4,5,6,7-tetrahydropyrrolo[2,3-c]pyridine-4,5,7-trione (28). To a solution of *N*-[(1-methylpyrrol-2-yl)carbonyl]-*N*-benzylglycine dimethylamide (26) (0.2995 g, 1.0 mmol) in acetonitrile (20 mL) at 0 °C was added phosphorus oxychloride (0.28 mL, 3.05 mmol, 3 equiv) and *N*-methylmorpholine (0.23 mL, 2.01 mmol, 2 equiv). After the addition was completed, the reaction mixture was heated at 65 °C for 21 h. The solution was cooled to room temperature and then concentrated under reduced pressure. The residue was dissolved in chloroform (20 mL), washed with saturated

aqueous Na2CO3 solution, saturated NaCl solution, dried over anhydrous MgSO4 and concentrated under reduced pressure. The first attempt to purify the resulting residue by flash column chromatography (Hex/EtOAc = 3:7, $R_f = 0.3$) gave a part of the crude product 27, which was characterized only by ¹H NMR. 6-Benzyl-4-dimethylamino-1-methylpyrrolo[2,3-*c*]pyridin-7-one (27): ¹H NMR (CDCl₃, 400 MHz) δ 2.70 (s, 6 H), 4.16 (s, 3 H), 5.16– 5.16 (m, 2 H), 6.29 (s, 1 H), 6.34 (d, 1 H, J = 2.9 Hz), 6.96 (d, 1 H, J = 2.8 Hz), 7.22-7.31 (m, 5 H). Further purification by flash column chromatography (Hex/EtOAc = 3:7, $R_f = 0.5$) afforded 28 (white amorphous solid, 0.1207 g, 0.45 mmol, 45%). The solid was recrystallized with toluene to give the analytical sample of 28 (white solid, 0.0982 g, 0.35 mmol, 37%). 6-Benzyl-1-methyl-4,5,6,7tetrahydropyrrolo[2,3-c]pyridine-4,5,7-trione (28): mp 151-154 °C (toluene); ¹H NMR (CDCl₃, 400 MHz) δ 4.04 (s, 3 H), 5.11 (s, 2 H), 6.71 (d, 1 H, J = 2.0 Hz), 6.92 (d, 1 H, J = 1.8 Hz), 7.23-7.31 (m, 3 H), 7.43–7.45 (m, 2 H); 13 C NMR (CDCl₃, 100 MHz) δ 36.7 (CH₃), 43.4 (CH₂), 108.2 (CH), 124.9, 127.4, 127.8 (CH), 128.5 (CH), 129.1 (CH), 132.1 (CH), 136.3, 157.1, 158.8, 169.7; MS (EI, 70 eV) m/z 107 (70), 108 (15), 177 (14), 240 (11), 268 (100) (M⁺); HRMS (EI, magnetic sector) calcd for C₁₅H₁₂N₂O₃ 268.0848, found 268.0851.

N-[(1-Methylpyrrol-2-yl)carbonyl]prolinol (5c). Method A. To a solution of N-[(1-methylpyrrol-2-yl)carbonyl]proline methyl ester (**5b**, 0.24 g, 1.02 mmol) in THF/EtOH (1/2 mL) was added LiCl (0.0861 g, 2.03 mmol, 2 equiv) and NaBH₄ (0.0768 g, 2.03 mmol, 2 equiv) at 0 °C. The reaction mixture was stirred for 8 h at room temperature then the solvent was removed under reduced pressure. The residue was dissolved in EtOAc (20 mL), washed with H₂O, saturated aqueous NaCl solution, dried over anhydrous Na₂SO₄, and then concentrated under reduced pressure. The residue was purified by flash column chromatography (Hex/EtOAc = 6:4, $R_f = 0.3$) to give **5c** (oil, 0.19 g, 0.92 mmol, 90%).

Method B. A mixture of 1-methyl-2-trichloroacetylpyrrole (0.23 g, 1.0 mmol), prolinol (151.6 mg, 1.50 mmol) and Na₂CO₃ (0.32 g, 3 mmol) in CH₃CN (10 mL) was heated at reflux temperature for 4 h. After cooling to room temperature, the solvent was removed under reduced pressure. The residue was dissolved in EtOAc (20 mL), washed with H2O, saturated aqueous NaCl solution, and then dried over anhydrous Na₂SO₄. The solvent was evaporated under reduced pressure. The residue was purified by flash column chromatography $(\text{Hex/EtOAc} = 6:4, R_f = 0.3)$ to give **5c** (oil, 0.19 g, 0.92 mmol, 92%): ¹H NMR (CDCl₃, 400 MHz) δ 1.58–1.67 (m, 1 H), 1.73–1.96 (m, 2 H), 2.08-2.16 (m, 1 H), 3.60-3.72 (m, 2 H), 3.84 (s, 3 H), 3.90-3.95 (m, 1 H), 4.41 (dd, 1 H, J = 2.7 and 7.5 Hz), 4.96 (bs, 1 H, OH), 6.09 (dd, 1 H, J = 2.7 and 3.7 Hz), 6.54 (dd, 1 H, J = 1.5 and 3.9 Hz), 6.70 (d, 1 H, J = 1.8 Hz); ¹³C NMR (CDCl₃, 100 MHz) δ 25.1 (CH₂), 28.3 (CH₂), 36.6 (CH), 50.9 (CH₂), 61.5 (CH), 67.6 (CH), 106.9 (CH), 114.3 (CH), 125.6, 127.2 (CH), 164.4; MS (FAB) m/z 84 (36), 108 (100), 209 (53) (M + 1); HRMS (FAB, magnetic sector) calcd for $C_{11}H_{17}N_2O_2$ 209.1290 (M + 1), found 209.1300.

1-Methyl-5,6,7,9-tetrahydropyrrolo[2,3-f]indolizin-9-one (29). A mixture of 5c (0.1000 g, 0.48 mmol) and IBX¹⁸ (0.4032 g, 1.44 mmol) in EtOAc (5 mL) was heated at reflux temperature for 3 h. After cooling to room temperature, the reaction mixture was filtered through Celite, and the solid was quickly washed with EtOAc. The combined filtrate was concentrated under reduced pressure to give the crude product of N-[(1-methylpyrrol-2-yl)carbonyl]-2-formylpyrrolidine (5d), which was used for subsequent reaction without further purification. To the solution of the crude 5d (0.48 mmol, obtained from the oxidation of 5c without purification) in CH_2Cl_2 (5 mL) was slowly added methanesulfonic acid (69.2 mg, 0.72 mmol, 1.5 equiv) at 0 °C. After the addition was completed, the reaction temperature was raised to room temperature, and the reaction mixture was stirred for 1.5 h under argon. The solvent was removed under reduced pressure. The residue was dissolved in EtOAc (25 mL), washed with H2O, saturated aqueous Na₂CO₃ solution, saturated aqueous NaCl solution, and then dried over anhydrous Na2SO4. The solvent was evaporated under reduced pressure. The residue was purified by flash column chromatography (Hex/EtOAc = 5:5, $R_f = 0.3$) to give 29 (amorphous

solid, 61.4 mg, 0.33 mmol, 68%). The solid was recrystallized with Hex/EtOAc to give the analytical sample of **29**: mp 89–91 °C (Hex/EtOAc); ¹H NMR (CDCl₃, 400 MHz) δ 2.11–2.20 (m, 2 H), 3.02 (t, 2 H, *J* = 7.6 Hz), 4.10 (t, 2 H, 7.2 Hz), 4.15 (s, 3 H), 6.15 (d, 1 H, *J* = 2.4 Hz), 6.32 (s, 1 H), 6.92 (d, 1 H, *J* = 2.4 Hz); ¹³C NMR (CDCl₃, 100 MHz) δ 22.7 (CH₂), 30.8 (CH₂), 35.6 (CH₃), 47.3 (CH₂), 96.1 (CH), 101.0 (CH), 121.2, 131.3, 132.7, 140.2, 155.0; MS (EI, 70 eV) *m*/*z* 187 (100), 188 (68) (M⁺); HRMS (EI, magnetic sector) calcd for C₁₁H₁₂ON₂ 188.0950, found 188.0944. Anal. Calcd for C₁₁H₁₂ON₂: C, 70.19; H, 6.43; N, 14.88. Found: C, 69.98; H, 6.36; N, 14.86.

ASSOCIATED CONTENT

S Supporting Information

¹H and ¹³C NMR spectra of all the synthesized compounds and X-ray structural data (CIF) of compound **28**. This material is available free of charge via the Internet at http://pubs.acs.org.

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Notes

The authors declare no competing financial interest.

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DEDICATION

Dedicated to Professor Hung-wen (Ben) Liu on the occasion of his 60th birthday.

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